Supplemental Table 1. Yeast strains used in this study.

Strain	Genotype*	Source
RSY10		[1]
RSY1696	cnc1::KANMX6	[2]
RSY1701	med13::HIS3	[3]
RSY1707	mid2::HIS3 mtl1::TRP1 wsc1::KANMX4	[4]
RSY1770	grr1 Δ ::his5 ⁺	[5]
RSY1726	cdk8Δ::KANMX4	[3]
RSY1949	gal83::KANMX4	This study
RSY2080	snf1::KANMX4	This study
YPDahl17	sak1::KANMX4	[6]
MML1445	sip1::natMX4 sip2:: KANMX4	[7]
MSY557	sip1::HIS3 sip2:: HIS3 gal83::HIS3	[8]
JGY1		[9]
JGY4	psk1::HIS3 psk2∆:: KANMX4	[10]
РЈ69-4	LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-lacZ gal4∆ gal80∆	[11]

Genotype of all strains is *MATa* ade2 ade6 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 except YPDahl17, MML1445, JGY1 and JGY4 which are *MATa* ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 and PJ69-4 which is *MATa* trp1-901 leu2-3,112 ura3-52 his3-200 gal4 Δ gal80 Δ .

Supplemental Table 2. Plasmids used in this study.

Plasmid Name	Gene	Epitope Tag	Marker	Promoter	2μ/ CEN	Reference
рВК38	CNC1	YFP	URA3	ADH1	CEN	[12]
рКС337	CNC1	CNC1	TRP1	ADH1	CEN	[13]
pKC801	MED13	3HA	URA3	TRP1	CEN	[5]
pLR166	CNC1 ^{5266A}	CNC1	TRP1	TRP1	CEN	[14]
рКС803	MED13	3HA	LEU2	ADH1	CEN	[5]
рКС805	MED13 ^{571-650deg∆}	3HA	URA3	ADH1	CEN	This study
pKC814	MED13 ^{742-844deg∆}	3HA	URA3	ADH1	CEN	This study
pDS8	GAL4AD-MED13 ⁵⁷¹⁻⁹⁰⁶	1HA	LEU2	ADH1	2μ	[5]
pDS15	GAL4AD-MED13 ⁵⁷¹⁻⁶⁵⁰	1HA	LEU2	ADH1	2μ	[5]
pDS16	GAL4AD-MED13 ⁶⁵¹⁻⁹⁰⁶	1HA	LEU2	ADH1	2μ	[5]
pDS32	GAL4AD-MED13 ⁷⁴²⁻⁸⁴⁴	1HA	LEU2	ADH1	2μ	[5]
pDS44	GAL4AD-MED13 ^{571-650 S636}	1HA	LEU2	ADH1	2μ	This study
pDS51	GAL4AD-MED13 ^{571-650 S636, S634A}	1HA	LEU2	ADH1	2μ	This study
pDS55	GAL4AD-MED13 ⁵⁷¹⁻⁶⁵⁰ 5587A	1HA	LEU2	ADH1	2μ	This study
pDS56	GAL4AD-MED13 ⁵⁷¹⁻⁶⁵⁰ S58A7, S636A, S634A	1HA	LEU2	ADH1	2μ	This study
pDS45	NLS-Med13 ¹⁻³⁰⁶	1HA	LEU2	ADH1	2μ	This study
pDS46	NLS-Med13 ³⁰⁶⁻⁵⁷⁰	1HA	LEU2	ADH1	2μ	This study
pDS47	NLS-Med13 ⁹⁰⁷⁻¹⁴²⁰	1HA	LEU2	ADH1	2μ	This study
pDS52	NLS-Med13 ³⁰⁷⁻⁵⁷⁰	1HA	LEU2	ADH1	2μ	This study
pDS54	GST-Med13 ^{571-650,S608A}	GST	AMP	-	-	This study
pJG1215	HIS ₆ -PSK1-KD	HIS ₆	AMP	-	-	[10]
pUM504	CDK8	1HA	TRP	GPD	CEN	[15]
pACT2	GAL4AD	1HA	LEU2	ADH1	2μ	[16]
pAS2	GAL4BD	1HA	TRP	ADH1	2μ	[16]
pAS2-Grr1	GAL4BD-GRR1	1HA	TRP	ADH1	2μ	[17]
pAS2- Grr1ΔLΔF	GAL4BD-GRR1Grr1∆L∆F	1HA	TRP	ADH1	2μ	[17]
pJG1465	GAL4AD-MED13 ⁵⁰⁴⁻⁷⁰³	1HA	TRP	ADH1	2μ	[9]
pAS2-	GAL4BD-PSK1	1HA	TRP	ADH1	2μ	[9]
Psk1						
JG1193	Snf1	8Мус	URA3	Snf1	CEN	[18]
JG1338	Snf1 ^{K84R}	8Myc	URA3	Snf1	CEN	[19]
	Snf1	8Myc	URA3	GAL1-10	2μ	[19]
	Snf1 ^{K84R}	8Myc	URA3	GAL1-10	2μ	[19]
pNLS-HA	SV40 NLS	1HA	LEU2	ADH1	2μ	This study
Mt-Cherry	Mito-targeting	mCherry	TRP1	ADH1	CEN	This study
pRS314	-	-	TRP1	-	CEN	[20]
pRS316	-	-	URA	-	CEN	[20]



FIGURE S1: Phosphorylation of cyclin C is not required for degradation of Med13⁵⁷¹⁻⁶⁵⁰. (A) $cnc1\Delta$ cells (RSY1696) harboring degron⁵⁷¹⁻⁶⁵⁰ (pDS15) and either wild-type cyclin C (pKC337) or a vector control (pRS314) were treated with 0.4 mM H₂O₂ for the timepoints indicated and Med13⁵⁷¹⁻⁶⁵⁰-HA levels analyzed by Western blot. Tub1 levels were used as loading controls. (B) Degradation kinetics of the degron⁵⁷¹⁻⁶⁵⁰ constructs shown in (A). Values represent averages ± SD from a total of at least two Western blots from independent experiments. (C) As in (A) except that $cnc1\Delta$ harboring either wild-type cyclin C (pKC337) or a phospho-mutant (cyclin C S266A, pLR166) was examined. (D) Degradation kinetics of results shown in (C).



FIGURE S2: The PAS kinase can associate with Med13 but is not required for its H_2O_2 mediated degradation. (A) Y2H analysis of cells harboring the Med13 construct shown with either Grr1 or Psk1. PJ69-4 cells harboring the Med13-activating domain constructs shown and either pAS-Grr1 or pAS2-Psk1 which has previously been shown to interact with Med13⁵⁰⁵⁻⁷⁰³ [9]. The cells were grown on *-LEU, -TRP* drop out medium to select for both plasmids (left panel) or *-TRP, -LEU, -HIS –ADE* (right panel) to test for Med13-Grr1 interaction. **(B)** Wild-type (RSY10) and *psk1*Δ *psk2*Δ (JGY4) cells harboring Med13-HA (pKC801) were treated with 0.4 mM H₂O₂ for the timepoints indicated and Med13 levels analyzed by Western blot. Tub1 levels were used as loading controls. **(C)** Degradation kinetics of the results shown in (B). Values represent averages ± SD from a total of at least two Western blots from independent experiments.



FIGURE S3: (A) Cells with the genotypes shown harboring Med13-HA (pKC801) were treated with 0.4 mM H_2O_2 for the timepoints indicated and Med13 levels analyzed by Western blot. Tub1 levels were used as loading controls. (B) Wild-type (RSY10) cells harboring either degron⁵⁷¹⁻⁶⁵⁰ (pDS15) or with the mutations shown were treated with 0.4 mM H_2O_2 for the timepoints indicated and analyzed by Western blot. Tub1 levels were used as loading controls. (C) Control experiment for the co-immunoprecipitation analysis shown in Fig. 5D showing that cells harboring a vector control are unable to pull down Snf1-myc whereas Cdk8-HA can.



FIGURE S4. Upper panel: cyclin C is released from the nucleus following H_2O_2 stress in *med13* Δ cells harboring Med13^{571-650deg Δ} HA (pKC805) as the only source of Med13. Fluorescence microscopy of mid-log phase *med13* Δ harboring pKC805 and cyclin C-YFP (pBK38) were analyzed as shown before and after 0.4 mM H_2O_2 stress. Cells were stained with Dapi to visualize the nucleus. Lower Panel: as in upper panel except that the mitochondrial marker mCherry was also expressed. Bar = 13 μ M.

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