

Effects of the intestinal microbiota on prostate cancer treatment by androgen deprivation therapy

Safae Terrisse¹, Laurence Zitvogel²⁻⁵ and Guido Kroemer^{6-8,*}

¹ Medical Oncology, Hôpital Saint-Louis, Paris, France.

² INSERM U1015, Equipe Labellisée - Ligue Nationale contre le Cancer, Villejuif, France.

³ University Paris Saclay, Gif-sur-Yvette, France.

⁴ Gustave Roussy, ClinicObiome, Villejuif, France.

⁵ Center of Clinical Investigations in Biotherapies of Cancer (CICBT) 1428, Villejuif, France.

⁶ Equipe labellisée par la Ligue contre le Cancer, Université de Paris Cité, Sorbonne Université, Institut Universitaire de France, Inserm U1138, Centre de Recherche des Cordeliers, Paris, France.

⁷ Metabolomics and Cell Biology Platforms, Gustave Roussy Comprehensive Cancer Institute, Villejuif, France.

⁸ Institut du Cancer Paris CARPEM, Department of Biology, Hôpital Européen Georges Pompidou, AP-HP, Paris, France.

* Corresponding Author:

Guido Kroemer, Equipe labellisée par la Ligue contre le Cancer, Université de Paris Cité, Sorbonne Université, Institut Universitaire de France, Inserm U1138, Centre de Recherche des Cordeliers, Paris, France; Email: kroemer@orange.fr

ABSTRACT Prostate cancer (PC) can be kept in check by androgen deprivation therapy (ADT, usually with the androgen synthesis inhibitor abiraterone acetate or the androgen receptor antagonist such as enzalutamide) until the tumor evolves to castration-resistant prostate cancer (CRPC). The transition of hormone-sensitive PC (HSPC) to CRPC has been explained by cancer cell-intrinsic resistance mechanisms. Recent data indicate that this transition is also marked by cancer cell-extrinsic mechanisms such as the failure of ADT-induced PC immunosurveillance, which depends on the presence of immunostimulatory bacteria in the gut. Moreover, intestinal bacteria that degrade drugs used for ADT, as well as bacteria that produce androgens, can interfere with the efficacy of ADT. Thus, specific bacteria in the gut serve as a source of testosterone, which accelerates prostate cancer progression, and men with CRPC exhibit an increased abundance of such bacteria with androgenic functions. In conclusion, the response of PC to ADT is profoundly influenced by the composition of the microbiota with its immunostimulatory, immunosuppressive and directly ADT-subversive elements.

INTRODUCTION

The development of different types of cancer is profoundly influenced by the microbiota, which can act locally to directly participate in oncogenesis (e.g., by activating cell-autonomous oncogenic processes or by eliciting pro-carcinogenic chronic inflammation) or affect the evolution of cancers at a distance, mostly by modulating the dialogue between malignant and immune cells in favor of immunosurveillance or its failure [1, 2]. Such long-distance effects are mediated by the intestinal microflora, which is the most abundant and diverse microbiota in the human body. In addition, the composition in the local and intestinal mi-

doi: 10.15698/mic2022.12.787

Received originally: 26.09.2022;

in revised form: 07.11.2022,

Accepted 09.11.2022,

Published 15.11.2022

Keywords: *Akkermansia muciniphila*, castration-resistant prostate cancer, hormoneotherapy, microbiome, *Ruminococcus gnavus*.

Abbreviations:

PC – prostate cancer, CRPC – castration-resistant prostate cancer, Treg – T CD4+FoxP3+ cells, Th17 – interleukin-17, LPS – lipopolysaccharide, HDC – histidine decarboxylase, ADT – androgen deprivation therapy, HSPC – hormone-sensitive PC, CRPC – castration-resistance PC, sjTRECcs – signal joint T-cell receptor excision circles, FMT – fecal microbiota transfer

crobiota, as well as its therapy-induced shifts, affect the response of cancers to chemotherapy, targeted therapy and immunotherapy [3, 4]. The connections between the microbiome and prostate cancer from carcinogenesis to response to therapies targeting the androgen pathway are also relevant. Prostate cancer (PC) is not an exception to this general rule, as we will summarize in this minireview.

CONTRIBUTION OF THE LOCAL MICROBIOTA TO PROSTATE CARCINO-GENESIS AND CANCER PROGRESSION

Benign prostatic hyperplasia and PC are associated with prostatitis, which may be caused by bacteria. Of note, the

most prevalent microorganism in the human prostate gland is *Cutibacterium* (formerly known as *Propionibacterium acnes*) [5], which may stimulate the infiltration of the tissue by regulatory T CD4+FoxP3+ cells (Treg) and T helper cell producing interleukin-17 (Th17) cells, hence causing local immunosuppression as well as procarcinogenic inflammation [6]. Thus, the local microbiota present in the genitourinary tract might contribute to carcinogenesis. In this context, it appears interesting that the presence of bacteria (and in particular anaerobic species from 5 genera: *Anaerococcus prevotii*, *Fenollaria sp. nov.*, *Fusobacterium nucleatum*, *Porphyromonas sp. nov.* or *asaccharolytica*, and *Peptoniphilus sp. nov.* or *harei*) in urine sediments strongly correlates with poor PC prognosis determined by the D'Amico score [7]. Altogether, the available evidence suggests that the local microbiota might affect prostate cancer progression (Fig. 1). This speculation appears particularly interesting in view of a recent report describing therapeutically relevant immunity against urothelium invasive *Escherichia coli* in the clinical response of bladder cancer patients to immunotherapy with pembrolizumab [8]. However, although *E. coli* is often identified in PC specimens [5], its contribution to procarcinogenic inflammation versus tumor-suppressive immunosurveillance has not yet been investigated.

CONTRIBUTION OF THE INTESTINAL MICROBIOTA TO PROSTATE CARCINOGENESIS AND CANCER PROGRESSION

Obesity is a major risk factor for PC development and progression [9], as well as a driver of intestinal dysbiosis, which can be defined as an imbalance in the microbes present in the microflora. Dysbiosis is often associated with an increase in gut permeability and endotoxemia, which is an increase in the circulating level of bacterial lipopolysaccharide (LPS) [10, 11]. In mice, dysbiosis induced by broad-

spectrum antibiotics favors the accumulation of LPS in subcutaneous and orthotopic PC, thereby activating local inflammation and promoting tumor growth [12]. High fat diet-induced dysbiosis also triggers an LPS-mediated pro-inflammatory pathway secondary to the LPS-induced up-regulation of the histamine-producing enzyme histidine decarboxylase (HDC) in mast cells infiltrating the tumor [13]. Although it is technically feasible to detect LPS in human tissue sections [14], we are not aware of any study investigating the presence of LPS in preneoplastic or malignant prostate specimens. Such studies are urgently awaited. Of note, in PC patients, the fecal abundance of *Proteobacteria* correlates with the presence of distant metastases [12]. Thus, there is circumstantial evidence suggesting that intestinal dysbiosis contributes to prostate carcinogenesis and tumor progression (Fig. 1).

POSITIVE IMPACT OF THE GUT MICROBIOTA ON THE EFFICACY OF ANDROGEN-DEPRIVATION THERAPY

There are several strategies for the clinical management of PC that range from the absence of immediate action ('wait and see') to surgical removal of the tumor, as well as from radiation therapy to aggressive chemotherapies [15]. In addition, most PC patients initially respond to androgen deprivation therapy (ADT, usually with the androgen synthesis inhibitor abiraterone acetate or the androgen receptor antagonist such as enzalutamide) before hormone-sensitive PC (HSPC) progresses to castration-resistance PC (CRPC) [16]. Although much emphasis has been laid on the cell autonomous response of PC cells to ADT and the consequent HSPC-CRPC transition, there is evidence that ADT also acts through PC cell non-autonomous mechanisms involving the immune system. Thus, in a mouse model of PC, the therapeutic response to ADT depends on the immune system, as indicated by the fact that antibody-mediated depletion of T cells or genetically determined

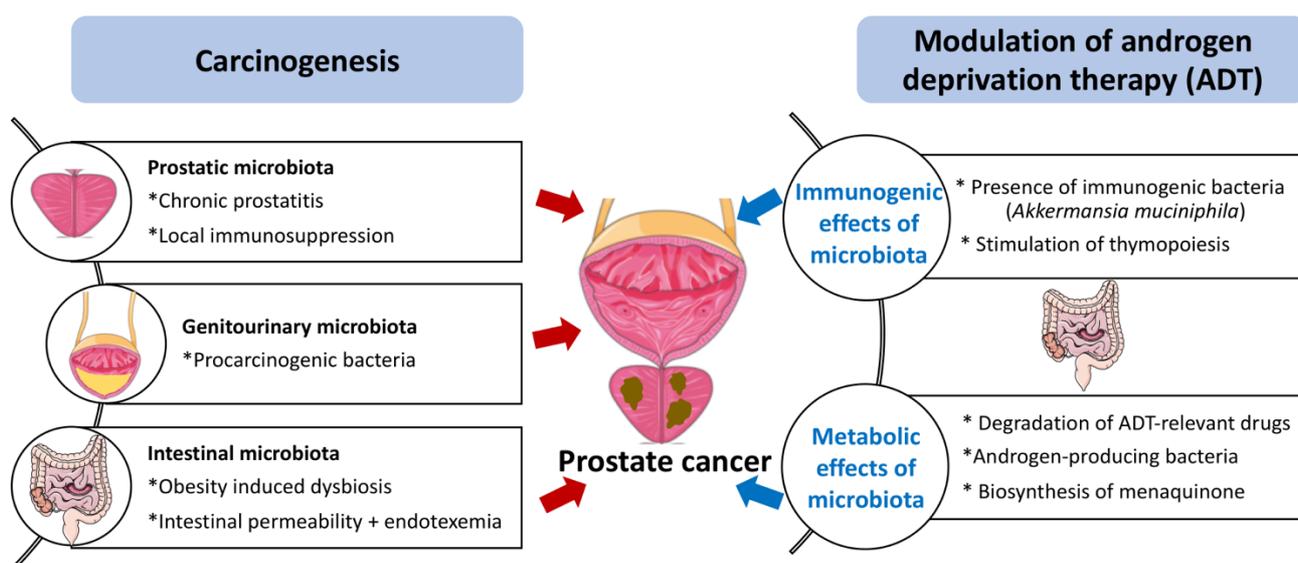


FIGURE 1: Potential mechanisms of action of the microbiota on prostate cancer carcinogenesis and sensitivity to androgen depletion therapy. For details consult text.

athymia compromise the ADT-mediated PC control [17]. This appears to be applicable to PC patients because successful (long-term) ADT results in an increase of thymic output, as indicated by the augmentation of recent thymic emigrant cells (i.e., signal joint T-cell receptor excision circles, abbreviated as sjTRECs) in peripheral blood [17].

As true for other cancers treated by immunogenic chemotherapy or immunotherapy [1, 2, 3, 4], the gut microbiota plays a major role in determining the therapy-relevant immune response elicited by ADT against PC. Thus, in the PC mouse model, depletion of the gut microbiota by orally administered broad-spectrum antibiotics reduces the efficacy of ADT. In mice, PC reduces the relative abundance of a particular immunostimulatory bacterium, *Akkermansia muciniphila*, in the gut, and this effect was reversed by ADT. Moreover, cohousing of PC-bearing mice with tumor-free mice or oral gavage with *A. muciniphila* ameliorates the efficacy of ADT. CRPC (but not HSPC) patients manifest a shift in the composition of their fecal microbiota that correlated with sjTRECs [17]. Although these results plead in favor of the beneficial impact of specific bacteria on the ADT-triggered anti-PC response. Nonetheless, the exact role of *A. muciniphila* in human PC remains controversial. For example, *A. muciniphila* might elicit both immune-dependent and immune-independent anticancer effects. Thus, in abiraterone-treated patients progressing towards CRPC, *A. muciniphila* expands, correlating with an increase in the biosynthesis of menaquinone (vitamin K2) [18] which inhibits PC growth in vitro, i.e., in an immune-independent fashion [19]. Conversely, extracellular vesicles derived from *A. muciniphila* can elicit cytotoxic T lymphocyte responses against PC in mice [20]. Moreover, according to one report, the clinical response to immunotherapy by PD-1 blockade of metastatic CRPC progressing on enzalutamide is associated with a decrease rather than with an increase in *A. muciniphila* [21], contrasting with the observation that *A. muciniphila* is associated with clinical responses to PD-1 blockade in other human malignancies including non-small cell lung cancer, melanoma and uroepithelial cancer [4, 22, 23]. Instead, the response of CRPC patients to PD-1 blockade correlated with an increase in the fecal abundance the *Streptococcus salivarius* [21]. Thus, different bacteria other than *A. muciniphila* may contribute to the clinical response of PC patients to immunotherapy. Irrespective of this detail, it appears clear that the gut microbiota contributes to the efficacy of ADT against PC (Fig. 1).

NEGATIVE EFFECTS OF THE GUT MICROBIOTA ON THE EFFICACY OF ANDROGEN-DEPRIVATION THERAPY

In sharp contrast with the afore-mentioned positive effects of the gut microbiota on ADT responses, there is also evidence supporting a deleterious role for specific bacteria in the therapeutic response. Thus, ADT of prostate cancer patients reportedly causes a decrease in α and β -diversity of the gut microbiota, which might precede or accompany the development of dysbiosis [24]. Prior reports indicate that specific bacteria contained in human feces can regulate androgen metabolism [25, 26, 27, 28]. More im-

portantly, however, it appears that bacteria can interfere with the pharmacokinetics and pharmacodynamics of ADT-relevant drugs.

On one hand, abiraterone can be metabolized by the gut microbiota in mice [29]. However, systematic clinical studies of the impact of intestinal bacteria on the half-life of abiraterone or other ADT-targeting molecules are elusive. On the other hand, perhaps more importantly, the gut microbiota from patients with CRPC or castrated mice can convert androgen precursors into active androgens, which are absorbed into the systemic circulation [30]. When the gut microbiota is depleted in mice, circulating DHEA and testosterone levels are significantly reduced. Furthermore, the *Ruminococcus* and *Bacteroides* genera are increased in the gut microbiota of CRPC patients compared to HSPC patients. In patients with CRPC, the *Ruminococcus* genus is associated with poor prognosis, while *Prevotella* is associated with favorable prognosis. *Ruminococcus gnavus* and *Bacteroides acidifaciens* can convert pregnenolone and hydroypregnenolone into androgens including DHEA and testosterone, and oral gavage with *R. gnavus* can indeed accelerate PC growth upon surgical castration. In contrast, the abundance of androgens is reduced and tumor growth is controlled upon fecal microbiota transfer (FMT) from hormone-sensitive prostate cancer patients or the administration of *Prevotella stercorea* into mice [30]. The mechanism through which *P. stercorea* reduces androgen production by the intestinal microbiota remains to be determined. Apart from this uncertainty, the available data plead in favor of the possibility that an androgen-producing microbiota subverts the impact of ADT against PC (Fig. 1).

CONCLUSION

As summarized in this review, the pathogenesis of PC, which - as true for other malignant diseases - involves chronic inflammation as well as failing immunosurveillance, may be conditioned by local infection and pro-inflammatory microbial products such as LPS. Intriguingly, it appears that the gut microbiota plays a decisive role in determining the fate of PC patients under ADT. Since ADT, at least in part, must induce an anticancer immune response to be efficient and the immune system is under the tonic influence of the intestinal microflora, dysbiosis and depletion of immunostimulatory bacteria may have a negative impact on the therapeutic efficacy of ADT. In addition, bacteria in the gut affect the metabolism of ADT-relevant drugs, reducing their concentrations in circulation, and produce steroids that interfere with hormone therapies (Fig. 1).

ACKNOWLEDGMENTS

GK and LZ are supported by the Ligue contre le Cancer (équipe labellisée); Agence National de la Recherche (ANR) – Projets blancs; Cancéropôle Ile-de-France; Fondation pour la Recherche Médicale (FRM); a donation by Elior; Equipex Onco-Pheno-Screen; Gustave Roussy Odyssey, the European Union Horizon 2020 Projects Oncobiome and Crimson; Institut National du Cancer (INCa); Institut Uni-

versitaire de France; LabEx Immuno-Oncology (ANR-18-IDEX-0001); a Cancer Research ASPIRE Award from the Mark Foundation; the RHU Immunolife; Seerave Foundation; SIRIC Stratified Oncology Cell DNA Repair and Tumor Immune Elimination (SOCRATE); and SIRIC Cancer Research and Personalized Medicine (CARPEM). This study contributes to the IdEx Université de Paris ANR-18-IDEX-0001.

AUTHOR CONTRIBUTION

GK wrote the first draft and then received major input from ST and LZ. All authors have read, edited and approved the paper.

CONFLICT OF INTEREST

GK has been holding research contracts with Daiichi Sankyo, Eleor, Kaleido, Lytix Pharma, PharmaMar, Osasuna Therapeutics, Samsara Therapeutics, Sanofi, Tollys, and Vascage. GK has been consulting for Reithera. GK is on the Board of Directors of the Bristol Myers Squibb Foundation France. GK is a scientific co-founder of everImmune, Osasuna Therapeutics, Samsara Therapeutics and Therfast Bio. GK is the inventor of patents covering therapeutic targeting of

aging, cancer, cystic fibrosis and metabolic disorders. GK's brother, Romano Kroemer, was an employee of Sanofi and now consults for Boehringer-Ingelheim. LZ has held research contracts with 9 Meters Biopharma, Daiichi Sankyo, Pilege, was on the on the Board of Directors of Transgene, is a cofounder of everImmune, and holds patents covering the treatment of cancer and the therapeutic manipulation of the microbiota.

COPYRIGHT

© 2022 Terrisse *et al.* This is an open-access article released under the terms of the Creative Commons Attribution (CC BY) license, which allows the unrestricted use, distribution, and reproduction in any medium, provided the original author and source are acknowledged.

Please cite this article as: Safae Terrisse, Laurence Zitvogel and Guido Kroemer (2022). Effects of the intestinal microbiota on prostate cancer treatment by androgen deprivation therapy. *Microbial Cell* 9(12): 190-194. doi: 10.15698/mic2022.12.787

REFERENCES

- Zitvogel L, Ma Y, Raoult D, Kroemer G, and Gajewski TF (2018). The microbiome in cancer immunotherapy: Diagnostic tools and therapeutic strategies. *Science*. 359(6382): 1366–1370. doi: 10.1126/science.aar6918
- Park EM, Chelvanambi M, Bhutiani N, Kroemer G, Zitvogel L, and Wargo JA (2022). Targeting the gut and tumor microbiota in cancer. *Nat Med*. 28(4): 690–703. doi: 10.1038/s41591-022-01779-2
- López-Otín C, and Kroemer G (2021). Hallmarks of health. *Cell*. 184(7): 1929–1939. doi: 10.1016/j.cell.2021.03.033
- Lee KA *et al.* (2022). Cross-cohort gut microbiome associations with immune checkpoint inhibitor response in advanced melanoma. *Nat Med*. 28(3): 535–544. doi: 10.1038/s41591-022-01695-5
- Brüggemann H, and Al-Zeer MA (2020). Bacterial signatures and their inflammatory potentials associated with prostate cancer. *APMIS*. 128(2): 80–91. doi: 10.1111/apm.13021
- Radej S, Szewc M, and Maciejewski R (2022). Prostate Infiltration by Treg and Th17 Cells as an Immune Response to Propionibacterium acnes Infection in the Course of Benign Prostatic Hyperplasia and Prostate Cancer. *Int J Mol Sci*. 23(16): 8849. doi: 10.3390/ijms23168849
- Hurst R, Meader E, Gihawi A, Rallapalli G, Clark J, Kay GL, Webb M, Manley K, Curley H, Walker H, Kumar R, Schmidt K, Crossman L, Eeles RA, Wedge DC, Lynch AG, Massie CE, CRUK-ICGC Prostate Group, Yazbek-Hanna M, Rochester M, Mills RD, Mithen RF, Traka MH, Ball RY, O'Grady J, Brewer DS, Wain J, and Cooper CS (2022). Microbiomes of Urine and the Prostate Are Linked to Human Prostate Cancer Risk Groups. *Eur Urol Oncol*. 5(4): 412–419. doi: 10.1016/j.euo.2022.03.006
- Goubet A-G *et al.* (2022). Escherichia coli-specific CXCL13-producing TFH are associated with clinical efficacy of neoadjuvant PD-1 blockade against muscle-invasive bladder cancer. *Cancer Discov*. CD-22-0201. doi: 10.1158/2159-8290.CD-22-0201
- Wilson RL, Taaffe DR, Newton RU, Hart NH, Lyons-Wall P, and Galvão DA (2022). Obesity and prostate cancer: A narrative review. *Crit Rev Oncol Hematol*. 169: 103543. doi: 10.1016/j.critrevonc.2021.103543
- Netto Candido TL, Bressan J, and Alfenas R de CG (2018). Dysbiosis and metabolic endotoxemia induced by high-fat diet. *Nutr Hosp*. 35(6): 1432–1440. doi: 10.20960/nh.1792
- Crocetto F, Boccellino M, Barone B, Di Zazzo E, Sciarra A, Galasso G, Settembre G, Quagliuolo L, Imbimbo C, Boffo S, Angelillo IF, and Di Domenico M (2020). The Crosstalk between Prostate Cancer and Microbiota Inflammation: Nutraceutical Products Are Useful to Balance This Interplay? *Nutrients*. 12(9): E2648. doi: 10.3390/nu12092648
- Zhong W, Wu K, Long Z, Zhou X, Zhong C, Wang S, Lai H, Guo Y, Lv D, Lu J, and Mao X (2022). Gut dysbiosis promotes prostate cancer progression and docetaxel resistance via activating NF-κB-IL6-STAT3 axis. *Microbiome*. 10(1): 94. doi: 10.1186/s40168-022-01289-w
- Matsushita M, Fujita K, Hatano K, Hayashi T, Kayama H, Motooka D, Hase H, Yamamoto A, Uemura T, Yamamichi G, Tomiyama E, Koh Y, Kato T, Kawashima A, Uemura M, Nojima S, Imamura R, Mubeen A, Netto GJ, Tsujikawa K, Nakamura S, Takeda K, Morii E, and Nonomura N (2022). High-fat diet promotes prostate cancer growth through histamine signaling. *Int J Cancer*. 151(4): 623–636. doi: 10.1002/ijc.34028
- Xia L, Xu Z, Zhou X, Bergmann F, Grabe N, Büchler MW, Neoptolemos JP, Hackert T, Kroemer G, and Fortunato F (2020). Impaired autophagy increases susceptibility to endotoxin-induced chronic pancreatitis. *Cell Death Dis*. 11(10): 889. doi: 10.1038/s41419-020-03050-3
- Rebello RJ, Oing C, Knudsen KE, Loeb S, Johnson DC, Reiter RE, Gillissen S, Van der Kwast T, and Bristow RG (2021). Prostate cancer. *Nat Rev Dis Primers*. 7(1): 9. doi: 10.1038/s41572-020-00243-0

16. Auchus RJ, and Sharifi N (2020). Sex Hormones and Prostate Cancer. *Annu Rev Med.* 71: 33–45. doi: 10.1146/annurev-med-051418-060357
17. Terrisse S *et al.* (2022). Immune system and intestinal microbiota determine efficacy of androgen deprivation therapy against prostate cancer. *J Immunother Cancer.* 10(3): e004191. doi: 10.1136/jitc-2021-004191
18. Daisley BA, Chanyi RM, Abdur-Rashid K, Al KF, Gibbons S, Chmiel JA, Wilcox H, Reid G, Anderson A, Dewar M, Nair SM, Chin J, and Burton JP (2020). Abiraterone acetate preferentially enriches for the gut commensal *Akkermansia muciniphila* in castrate-resistant prostate cancer patients. *Nat Commun.* 11(1): 4822. doi: 10.1038/s41467-020-18649-5
19. Dasari S, Samy ALPA, Kajdacsy-Balla A, Bosland MC, and Muni-rathinam G (2018). Vitamin K2, a menaquinone present in dairy products targets castration-resistant prostate cancer cell-line by activating apoptosis signaling. *Food Chem Toxicol.* 115: 218–227. doi: 10.1016/j.fct.2018.02.018
20. Luo Z-W, Xia K, Liu Y-W, Liu J-H, Rao S-S, Hu X-K, Chen C-Y, Xu R, Wang Z-X, and Xie H (2021). Extracellular Vesicles from *Akkermansia muciniphila* Elicit Antitumor Immunity Against Prostate Cancer via Modulation of CD8+ T Cells and Macrophages. *Int J Nanomedicine.* 16: 2949–2963. doi: 10.2147/IJN.S304515
21. Peiffer LB, White JR, Jones CB, Slottke RE, Ernst SE, Moran AE, Graff JN, and Sfanos KS (2022). Composition of gastrointestinal microbiota in association with treatment response in individuals with metastatic castrate resistant prostate cancer progressing on enzalutamide and initiating treatment with anti-PD-1 (pembrolizumab). *Neoplasia.* 32: 100822. doi: 10.1016/j.neo.2022.100822
22. Routy B *et al.* (2018). Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science.* 359(6371): 91–97. doi: 10.1126/science.aan3706
23. Derosa L *et al.* (2022). Intestinal *Akkermansia muciniphila* predicts clinical response to PD-1 blockade in patients with advanced non-small-cell lung cancer. *Nat Med.* 28(2): 315–324. doi: 10.1038/s41591-021-01655-5
24. Kure A, Tsukimi T, Ishii C, Aw W, Obana N, Nakato G, Hirayama A, Kawano H, China T, Shimizu F, Nagata M, Isotani S, Muto S, Horie S, and Fukuda S (2022). Gut environment changes due to androgen deprivation therapy in patients with prostate cancer. *Prostate Cancer Prostatic Dis.* doi: 10.1038/s41391-022-00536-3
25. Collén H, Landin A, Wallenius V, Elebring E, Fändriks L, Nilsson ME, Ryberg H, Poutanen M, Sjögren K, Vandenput L, and Ohlsson C (2019). The gut microbiota is a major regulator of androgen metabolism in intestinal contents. *Am J Physiol Endocrinol Metab.* 317(6): E1182–E1192. doi: 10.1152/ajpendo.00338.2019
26. Ly LK, Rowles JL, Paul HM, Alves JMP, Yemm C, Wolf PM, Devendran S, Hudson ME, Morris DJ, Erdman JW, and Ridlon JM (2020). Bacterial steroid-17,20-desmolase is a taxonomically rare enzymatic pathway that converts prednisone to 1,4-androstenedione-3,11,17-trione, a metabolite that causes proliferation of prostate cancer cells. *J Steroid Biochem Mol Biol.* 199: 105567. doi: 10.1016/j.jsbmb.2019.105567
27. Devendran S, Mythen SM, and Ridlon JM (2018). The *desA* and *desB* genes from *Clostridium scindens* ATCC 35704 encode steroid-17,20-desmolase. *J Lipid Res.* 59(6): 1005–1014. doi: 10.1194/jlr.M083949
28. Ridlon JM, Ikegawa S, Alves JMP, Zhou B, Kobayashi A, Iida T, Mitamura K, Tanabe G, Serrano M, De Guzman A, Cooper P, Buck GA, and Hylemon PB (2013). *Clostridium scindens*: a human gut microbe with a high potential to convert glucocorticoids into androgens. *J Lipid Res.* 54(9): 2437–2449. doi: 10.1194/jlr.M038869
29. Keranmu A, Yang F-Y, Wahafu W, Han S-J, Yang G-S, and Xing N-Z (2022). Biotransformation of Abiraterone Into Five Characteristic Metabolites by the Rat Gut Microbiota and Liver Microsomes. *Front Oncol.* 12: 890323. doi: 10.3389/fonc.2022.890323
30. Pernigoni N *et al.* (2021). Commensal bacteria promote endocrine resistance in prostate cancer through androgen biosynthesis. *Science.* 374(6564): 216–224. doi: 10.1126/science.abf8403