Supplementary Information for Exploring Carbon Source related Localization and Phosphorylation in the Snf1/Mig1 Network using Population- and Single Cell-based Approaches

Svenja Braam¹, Farida Tripodi^{2 **,} Linnea Österberg^{1,3}, Sebastian Persson¹, Niek Welkenhuysen^{1,3}, Paola Coccettⁱ², Marija Cvijovic^{1*}

 Department of Mathematical Sciences, Chalmers University of Technology and University of Gothenburg, Sweden
 Department of Biotechnology and Biosciences, University of Milano-Bicocca, Italy

3 Department of Biology and Biological Engineering, Chalmers University of Technology, Sweden

*Corresponding author: Marija Cvijovic Department of Mathematical Sciences, Chalmers University of Technology and University of Gothenburg SE-412 96 Gothenburg Sweden

marija.cvijovic@chalmers.se +46317725321

** Co-corresponding author:
Farida Tripodi
Department of Biotechnology and Biosciences
University of Milano-Bicocca
Piazza della Scienza 2, 20126 Milano
Italy
farida.tripodi1@unimib.it
+39 264483513



Figure S1, Supplementary data:

Mig1 nuclear localization ratio in cells 15 minutes after the shift from ethanol to the indicated concentration of glucose, fructose or mannose. Horizontal black lines indicate the median.



Figure S2, Supplementary data:

A) Mean of relative Snf1 phosphorylation by western blot quantification 5, 15 and 30 minutes after upshift from ethanol to glucose, fructose or mannose, dots represent biological replicates. B) Western blot images of all replicates tested for Snf1 phosphorylation quantification. Fru = fructose, Glc = glucose, Man = mannose Exemplary images also found in Figure 2C.



Figure S3, Supplementary data:

Three individual colonies of the indicated strains of the BY4741 background were grown over night in YPD supplemented with 2% glucose. The medium was removed, cells were washed once and diluted to an OD/ml of 0.1 in sterile water. Drops of 5 μ l of OD/ml 0.1 and ten fold dilutions up to 10⁻⁵ were spotted onto YPD agar plates with respective carbon source and grown at 30°C for three days before imaging.



Figure S4, Supplementary data:

Measurements of cell- and nucleus area of cells subjected to FRAP.

Violin plots show the distribution as well as boxplot for the measured data. Horizontal lines indicate the mean, the boxplot has as lower and upper hinge respectively the 25th and 75th percentile and the whiskers denote the 95% confidence interval. Black dots denote outliers.

S2: Monolix report: 0.05% glucose

Run: 240213_glu005_corr.mlxtran Dataset: 240213_FRAP_complete_norm_one_nominus_glu005_corr.csv Date: 02-13-2024

Tables

Darameter	Deremeter Value		STOCH. APPROX.	
Parameter Value		aiue	S.E.	R.S.E.(%)
Fixed Effects				
I0_pop	0.152		0.0054	3.554
A1_pop	0.168		0.0103	6.136
tau_pop	0.0638		0.00839	13.146
Standard Deviation of the Random Effects				
	Value	C.V.(%)		
omega_l0	0.16	16.151	0.026	16.208
omega_A1	0.279	28.486	0.0466	16.68
omega_tau	0.566	61.425	0.1	17.684
Error Model Parameters				
b	0.135		0.00207	1.529

Table 1: Estimated population parameters

Table 2: Log-likelihood and Information criteria

CRITERIA	IMPORTANCE SAMPLING
-2 x log-likelihood (OFV)	-8675.2
Akaike Information Criteria (AIC)	-8661.2
Bayesian Information Criteria (BIC)	-8653
Corrected Bayesian Information Criteria (BICc)	-8634.8

Plots



Figure 1: Observed data



Figure 2: Individual fits



Figure 3: Observations vs predictions



Figure 4: Scatter plot of the residuals



Figure 5: Distribution of the residuals



Figure 6: Visual predictive check



Figure 7: Distribution of the individual parameters



Figure 8: Distribution of the standardized random effects



Figure 9: Correlation between random effects



Figure 10: SAEM

S2: Monolix report: 2% glucose

Run: 240213_glu2_corr.mlxtran Dataset: 240213_FRAP_complete_norm_one_nominus_glu2_corr.csv Date: 02-13-2024

Tables

Table 5. Estimated population parameters				
Parameter Value		STOCH. APPROX.		
		alue	S.E.	R.S.E.(%)
Fixed Effects				
I0_pop	0.156		0.00607	3.888
A1_pop	0.172		0.011	6.38
tau_pop	0.0666		0.0074	11.123
Standard Deviation of the Random Effects				
	Value	C.V.(%)		
omega_l0	0.184	18.542	0.0286	15.529
omega_A1	0.302	30.925	0.0474	15.684
omega_tau	0.491	52.16	0.0827	16.868
Error Model Parameters				
b	0.128		0.00193	1.503

Table 3: Estimated population parameters

Table 4: Log-likelihood and Information criteria

CRITERIA	IMPORTANCE SAMPLING
-2 x log-likelihood (OFV)	-9120.5
Akaike Information Criteria (AIC)	-9106.5
Bayesian Information Criteria (BIC)	-9098
Corrected Bayesian Information Criteria (BICc)	-9079.8

Plots



Figure 11: Observed data



time



Figure 12: Individual fits

time



Figure 13: Observations vs predictions



Figure 14: Scatter plot of the residuals



Figure 15: Distribution of the residuals



Figure 16: Visual predictive check



Figure 17: Distribution of the individual parameters



Figure 18: Distribution of the standardized random effects



Figure 19: Correlation between random effects



Figure 20: SAEM

S2: Monolix report: 2%Glycerol

Run: 240213_gly2_corr.mlxtran

Dataset: 240219_FRAP_complete_norm_one_nominus_gly2_corr.csv **Date:** 02-19-2024

Tables

ruble 5. Estimated population parameters				
Darameter	ator Value		STOCH. APPROX.	
Parameter Value		S.E.	R.S.E.(%)	
Fixed Effects				
I0_pop	0.152		0.00646	4.239
A1_pop	0.157		0.00892	5.674
tau_pop	0.0774		0.0124	15.977
Standard Deviation of the Random Effects				
	Value	C.V.(%)		
omega_l0	0.185	18.621	0.0327	17.725
omega_A1	0.243	24.661	0.0411	16.929
omega_tau	0.685	77.393	0.122	17.8
Error Model Parameters				
b	0.124		0.00201	1.626

Table 5: Estimated population parameters

Table 6: Log-likelihood and Information criteria

CRITERIA	IMPORTANCE SAMPLING
-2 x log-likelihood (OFV)	-7940.2
Akaike Information Criteria (AIC)	-7926.2
Bayesian Information Criteria (BIC)	-7918.9
Corrected Bayesian Information Criteria (BICc)	-7900.7

Plots



Figure 21: Observed data



time

Figure 22: Individual fits



Figure 23: Observations vs predictions



Figure 24: Scatter plot of the residuals



Figure 25: Distribution of the residuals



Figure 26: Visual predictive check



Figure 27: Distribution of the individual parameters



Figure 28: Distribution of the standardized random effects



Figure 29: Correlation between random effects



Figure 30: SAEM