

Supplemental material

Specific mutations in the permease domain of septal protein SepJ differentially affect functions related to multicellularity in the filamentous cyanobacterium *Anabaena*

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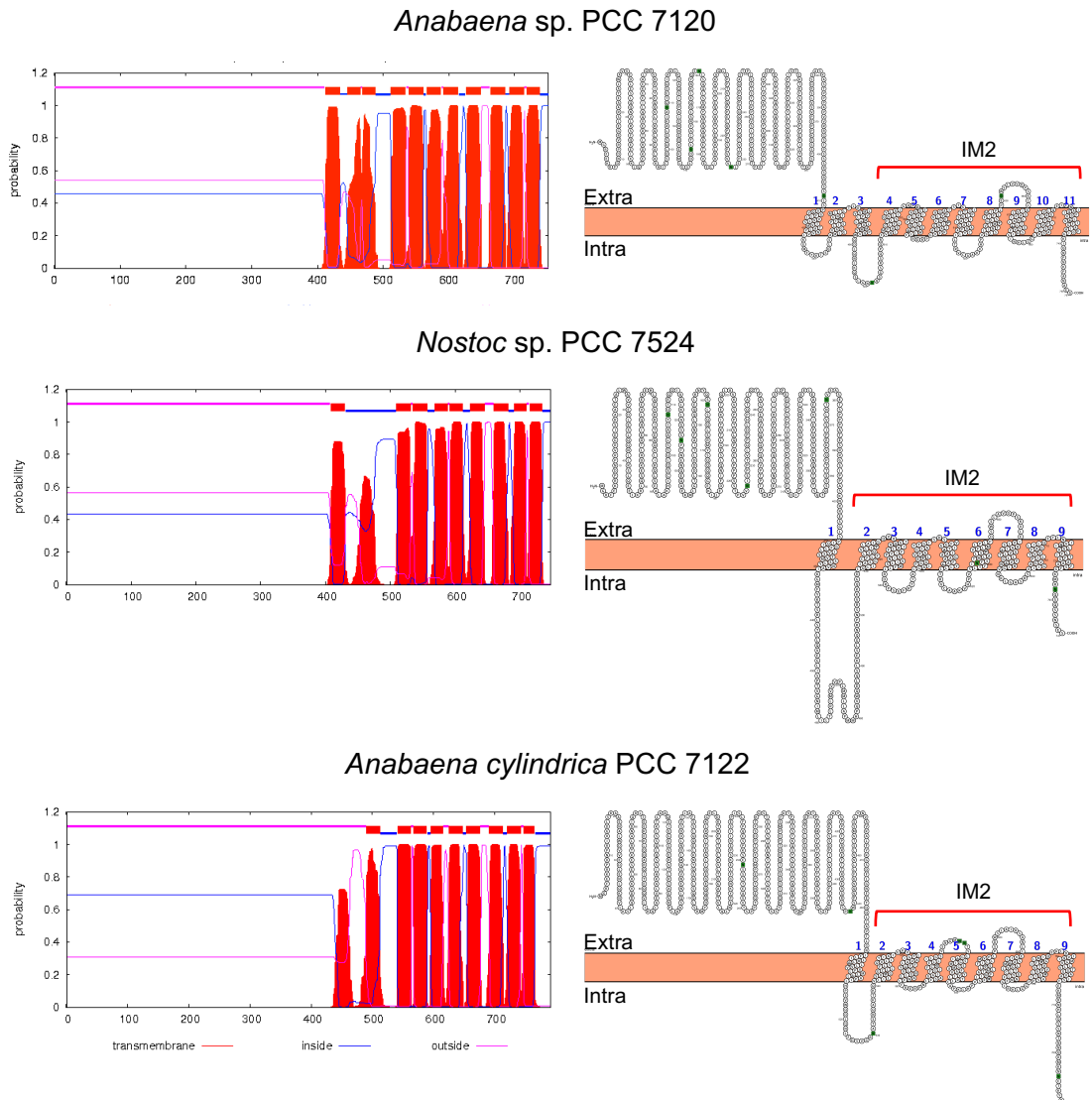


Fig. S1. Predicted topology of SepJ from *Anabaena* sp. PCC 7120 and, chosen as two further examples, *Nostoc* sp. PCC 7524 and *Anabaena cylindrica* PCC 7122. Left panels, TMHMM predictions; right panels, Protter representations of the topological features predicted in the left panels. These topologies illustrate the strong conservation of the last eight transmembrane segments (integral membrane protein subdomain 2, IM2 [ref. 1]), which is widely found for SepJ from 20 heterocyst-forming cyanobacteria that were inspected. N-terminal to this section of SepJ, two or three hydrophobic regions, which different topology programs identify as one, two or three possible TMSs, are found. When two TMSs are predicted (50% of the checked sequences), the N-terminal extra-membrane section is predicted to be cytoplasmic rather than periplasmic. As noted in the main text, available experimental evidence favors a periplasmic location of the N-terminal extra-membrane section of SepJ. Extra, extra-cytoplasmic space; Intra, cytoplasm.

A

>SepJ($\Delta 463-748$)

MGRFEKRPDNDPRVREGELSRAAETALWAVVEDLESLQQNVLRSFQEEIKKLQ
TEKDRLTDEVQQLIEEKEHLQEVRRITEQQVLIRQLSEALAKHICSQLQSSLAKI
ANQTESQIAALKSAQSIGPAIENNEQVEKMLGSLDDNLTIAFNSLQQELKNYQS
NLSQQLSRMYNQQQQGETIVEELIDRLRGELTRAIQETSTAKAQLSPPTVLQP
PELQPPSSPVVNLSPPTVLQFPDQQSPNPLQASTPLEETSTTKPSVSIPTPEK
STPVTIVPPPQETRPETKSVIPKVSPDSETKLQSSQEKAAPSSVINRELSAGA
AKSPLTPEKPPPEPISTSKTKFSPSSEKPPPEPISTSKTKFSPSSEKPPPEVSVLSR
DSSASKASTPPPAPVVRGSTPSSSRSRKSSNLSPVQVGFLLVVLSTVMTALY
NVVLKGMFYKTSQLSAMLEVAGLISPTLGNIMNSNAEG

B

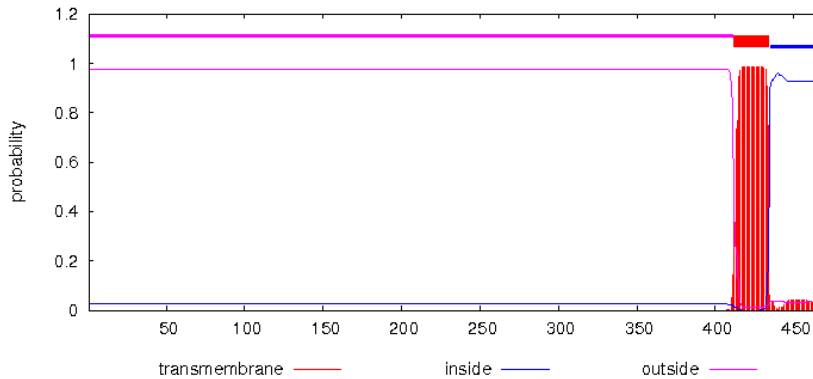


Fig. S2. The SepJ protein produced in strain CSVM90 (SepJ $\Delta 463-748$). (A) Amino acid sequence of SepJ $\Delta 463-748$. Red color, amino acids of the coiled-coil domain; green color, liker domain. The TMS starts at the VQVG sequence. A three-amino-acid-residue insertion (NSN) is present in this protein as a result of the construction procedure. (B) TMHMM prediction of the only transmembrane segment present in the SepJ $\Delta 463-748$ protein. The program also predicts that the N-terminal section of the protein is periplasmic.

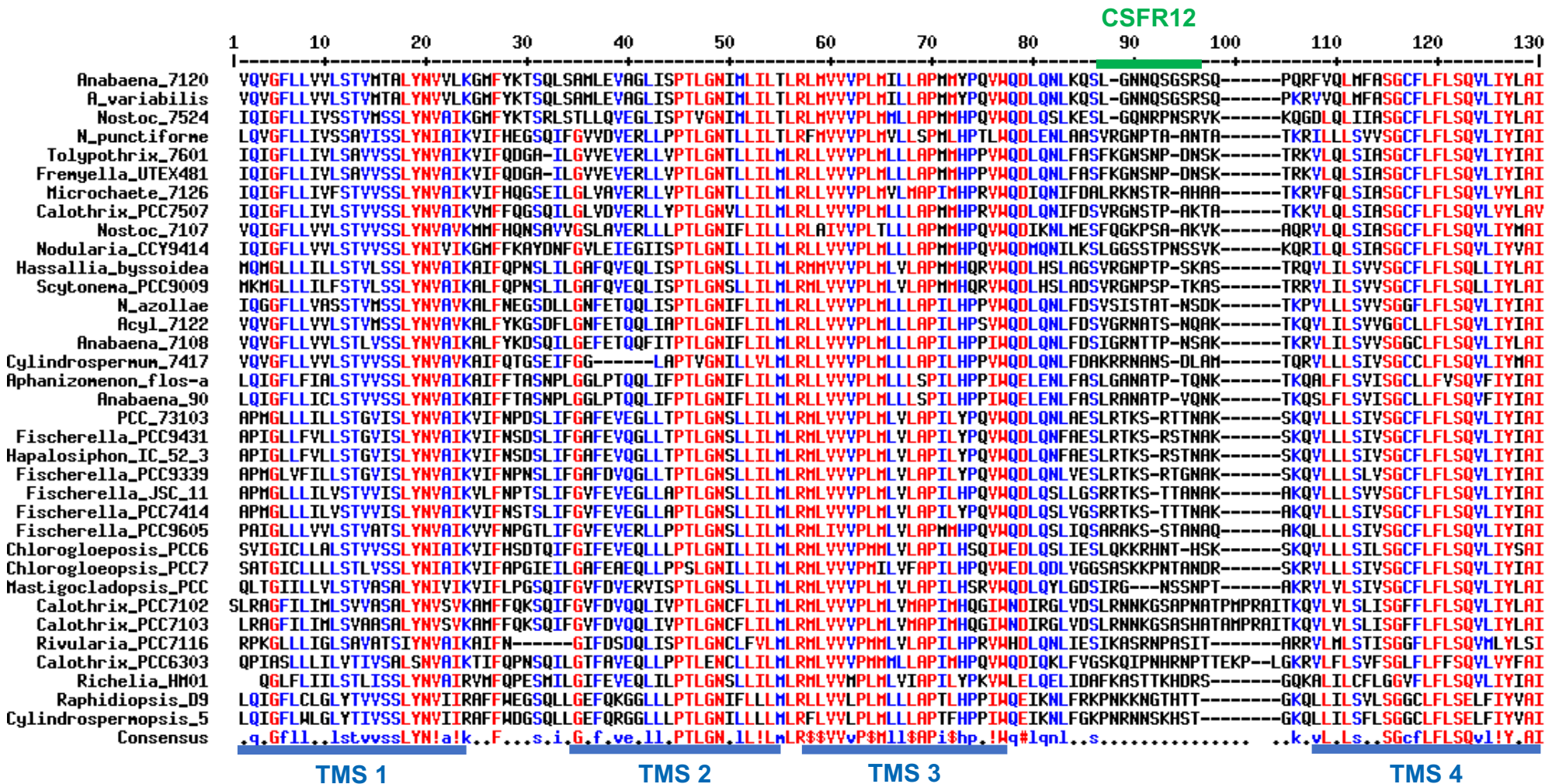


Fig. S3. Alignment of the permease section of SepJ from 35 heterocyst-forming cyanobacteria and location of the mutations introduced in the protein from *Anabaena* sp. strain PCC 7120. The sequence of strain PCC 7120 SepJ includes amino acid residues 412 to 751. The alignment was performed at MultAlin (<http://multalin.toulouse.inra.fr/multalin/>) with default parameters. Color code: red, amino acid residues conserved in more than 90% of the sequences; blue, amino acid residues conserved in more than 50% but less than 90% of the sequences. Horizontal green line, part of a predicted cytoplasmic loop in SepJ of heterocyst-forming cyanobacteria, deleted in strain CSFR12; blue vertical arrows, point mutations in the indicated strains; purple vertical arrow, residue at which the C-terminal deletion starts in strain CSFR22. Horizontal blue lines, TMSs as predicted by TMHMM for the SepJ protein of strain PCC 7120 (see Fig. S1).

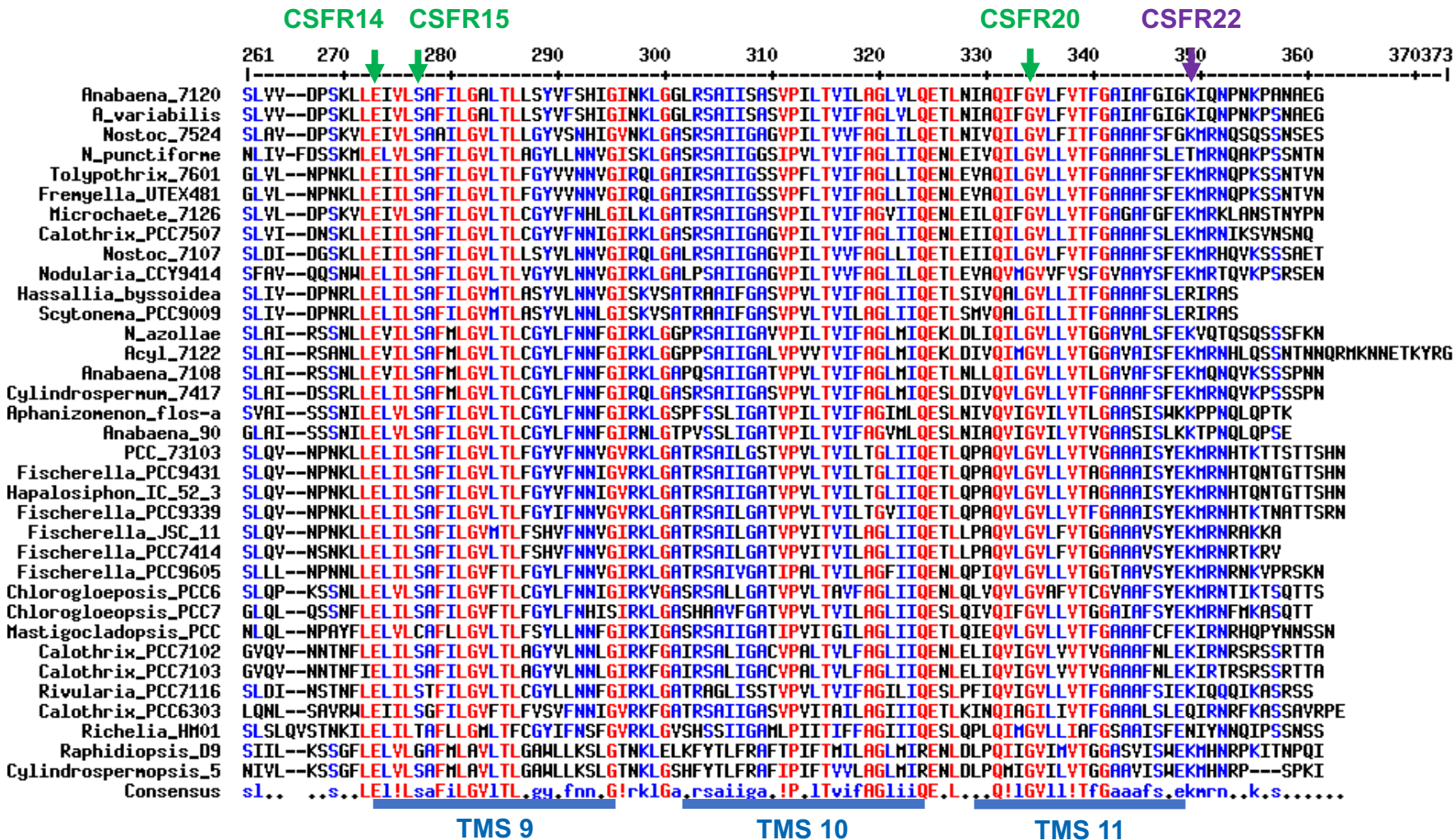


Fig. S3. Continued.

The SepJ proteins aligned are from the following cyanobacteria: *Anabaena* sp. PCC 7120, *Anabaena variabilis* ATCC29413, *Nostoc* sp. PCC 7524, *Nostoc punctiforme* PCC 73102, *Tolypothrix* sp. PCC 7601, *Fremyella diplosiphon* UTEX 481, *Microchaete* sp. PCC 7126, *Calothrix* sp. PCC 7507, *Nostoc* sp. PCC 7107, *Nodularia spumigena* CCY9414, *Hassallia byssoida*, *Scytonema* sp. PCC 9009, *Nostoc* (*Anabaena*) *azollae*, *Anabaena cylindrica* PCC 7122, *Anabaena* sp. PCC 7108, *Cyndrospermum stagnale* PCC 7417, *Aphanizomenon flos-aquae*, *Anabaena* sp. 90, *Fischerella muscicola* PCC 73103, *Fischerella* sp. PCC 9431, *Hapalosiphon* sp. IC-52-3, *Fischerella* sp. PCC 9339, *Fischerella* sp. JSC-11, *Fischerella* sp. PCC 7414, *Fischerella* sp. PCC 9605, *Chlorogloeopsis* sp. PCC 6912, *Chlorogloeopsis* sp. PCC 7702, *Mastigocladopsis* sp. PCC 10914, *Calothrix* sp. PCC 7102, *Calothrix* sp. PCC 7103, *Rivularia* sp. PCC 7116, *Calothrix* sp. PCC 6303, *Richelia intracellularis* HH01, *Raphidiopsis brookii* sp. D9, *Cyndrospermopsis* sp. 509.

Table S1. Oligodeoxynucleotide primers used in this work.

Primer	Sequence (5' → 3') ^a
alr2338-10	TGAGCCAGAAGTCCAGAG
alr2338-25	<u>CTGCAGAATCCTTTACAAGC</u>
alr2338-29	<u>GAATTCCAATGCAGAAGGTTA</u>
alr2338-30	<u>GAATTCATAATATTTCCCAAGGTC</u>
alr2338-34	<u>ACTAGTCCGGAACCTTATCTGC</u>
alr2338-37	ATCAG <u>AATCCC</u> ATTAGAGGAGACTAGC
alr2338-38	TAC <u>GAATTC</u> ATAGAGCGTTCTTATCTGC
alr2338-39	TGAAGCAATCACAGCCACAGAGATTTG
alr2338-40	TGGCTGTGATTGCTTCAGGTTTTGC
alr2338-41	ATCCTGACCGCCGTATGTGC
alr2338-42	ACATACGGCGGTCAGGATGAG
alr2338-43	TAAGTTACTGGCAATGTGTTAAGTGC
alr2338-44	GCACTTAACACAATTGCCAGTAACTTAG
alr2338-45	GAAATTGTGTTAGCTGCTTTTATTTTGGG
alr2338-46	CAAATAAAAAGCAGCTAACACAATTTCCAG
alr2338-47	GGCTAAAGTCGCTCCAGTATCTTTGTC
alr2338-48	AAAGATACTGGAGCGACTTTAGCCGCAC
alr2338-49	ATTTTCTGTGCTGAGGTGCTAGTTTTTG
alr2338-50	TAGCACCTCAGCACAGAAAATAGCG
alr2338-51	CAAATATTCGCAGTTTTATTCGTCACTTTTG
alr2338-52	GAATAAAACTGCGAATATTTGGGCAATG
alr2338-53	ACCGGAGTGCGTATCGCCCTTTTC
alr2338-54	AAGGGCGATACGCACTCCGGTGGG
alr2338-55	TAC <u>GAATTC</u> TTAGCCAATACCAAAAGC
alr2338-58	AGGATGAGGGCACAAAGCAAAGG
alr2338-59	TGCTTGTGCCCTCATCTGAC
alr2338-60	GCGGGCCAGGATGAGG
alr2338-61	ATCCTGGCCCGCGTATGTG
alr2338-62	GGGCGATCAGCGAACAGAAGCCAG
alr2338-63	TTCGCTGATCGCCCCGGTGTATTTTC
alr2338-64	TGTGGTGCAGGTGCTAGTTTTTG
alr2338-65	TAGCACCGCACACAGAAAATAGC

^a Restriction sites are underlined, and sequences modified for site-specific mutagenesis are shown in bold.

Table S2. Plasmids and oligonucleotides used for strain construction

Strain	Conjugated plasmid	Primer “a”	Primer “b”	SepJ version expressed
CSFR11	pCSFR53			wild type
CSFR12	pCSFR54	alr2338-40	alr2338-39	Δ (L498-S507)
CSFR13	pCSFR55	alr2338-42	alr2338-41	R617A
CSFR14	pCSFR56	alr2338-44	alr2338-43	E663A
CSFR15	pCSFR57	alr2338-46	alr2338-45	S667A
CSFR16	pCSFR58	alr2338-48	alr2338-47	H624A
CSFR19	pCSFR59	alr2338-50	alr2338-49	G579A
CSFR20	pCSFR60	alr2338-52	alr2338-51	G724A
CSFR21	pCSFR61	alr2338-54	alr2338-53	A542R
CSFR22	pCSFR62		alr2338-55	Δ (K739-G751)
CSFR25	pCSFR65	alr2338-58	alr2338-59	Y612A
CSFR26	pCSFR66	alr2338-60	alr2338-61	T616A
CSFR27	pCSFR67	alr2338-62	alr3838-63	R562A
CSFR28	pCSFR68	alr2338-64	alr2338-65	E580A